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(54) METHOD FOR DETERMINING FRAMESHIFT MUTATIONS IN CODING NUCLEIC ACIDS

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(52) U.S. Cl.

C12N 15/67

CPC C12Q 1/6827 (2013.01); C12N 15/63 (2013.01); C12N 15/67 (2013.01); C12Q 1/6897 (2013.01)

(2006.01)

(58) Field of Classification Search

See application file for complete search history.

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Primary Examiner — Jennifer Dunston

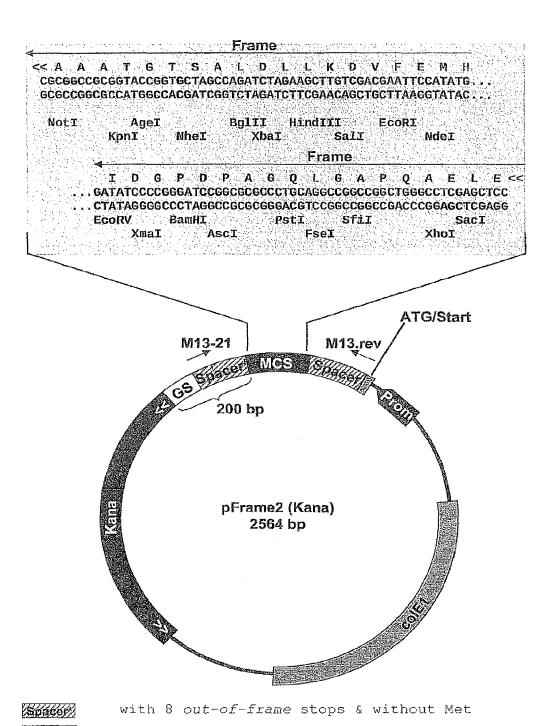
(57)ABSTRACT

The present invention relates to a method for identifying frameshift mutations in coding nucleic acid sequences.

10 Claims, 11 Drawing Sheets

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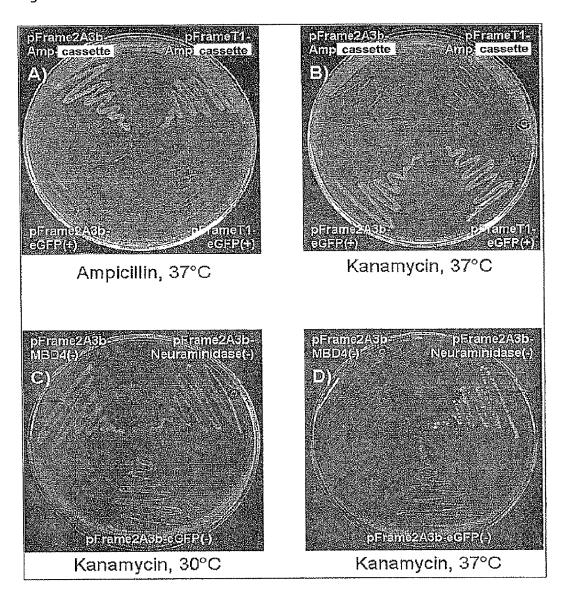
GS



glycine-serine linker

Figure 1

Figure 2



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Figure 3

A)	eGFP(-)	sequence	map	(reverse	complement)

Top: translation of minus strand

rop:	translation of minus strand	
Botto	m: translation of plus strand (wt protein)	
1 1	E L K L V Q F I H A Q G N T S G G H E F Q Q H H V I T GAGGTCAAGGTTGATGCATGCCAGGGTAATACCAGGGGGGGTGAGAATTCCAGGAGCACCATGTGATCAC CTCGAGTTCGAACATGTCAACATGTACGGTACG	80 89
81 81	F F V R I F A Q G A L G A Q V V V I R Q Q H R A I T GTTTTTCGTTCGGATCTTTGCTCAGGGCGCTCTGGGTGCTCAGGTAGTGGTTATCGGCAGCAGCACCACCAGCAGCAGCAGCAGCAGCAGCAGC	160 160
161 161	N R G V L L V V I G Q L H A A I F D V V A D F E V H F ATCEGEGETTTTC5CTGETAGTCATCCAGCTGCCAGCTGCCATCTTCGATGTTGTGGCGGATTTTGAAGTTCACTTTTAGCCCCACCAGAGACGACCATCACTAGCCGGTCGACGTGCACGGTAGAAGCTACAACACCGCCTAAAACTTCAAGTGAAA I P T N Q Q Y H D A L Q V S G D E I N H R I K F N V K	240 240
24 <u>1</u> 241	D A V F L F I G H D V H V V A V V V V F Q F V A Q D V GATGCCGTTTTTCTGTTTATCGGCCATGATGTACACGTTGTGGCCCAGGATGT CTACGGCAAAAAGACAAATAGCCGGTACTACATGTGCAACACCGACAACATCAACATAAGGTCAAACACCGGGTCCTACA I G N K Q K D A M I Y V N H S N Y N Y E L K H G L I N	320 320
321 321	T V L F K V D A F Q F D T V H Q G I A F E F H F G T TACCGTCCTCTTTAAAGTCGATECCTTTCAGTTCGATACGGTTCACCAGGGTATCCCCTTCGAATTCACTTCGGCAGGGAGAGAAATTTCAGCTACGGAAAGTCAAGCTATGCCAAGTGGTCCCATAGCGGAAGCTTAAAGTGAAGCCGTGCC G D E K F D I G K L E I R N V L T D G E F K V E A R	400 400
401 401	G F V V A I I F E E N G T F L H V A F R H G A F E E I GTTTTGTAGTTGCCATCATCATCTTTGAAGAAATC CAAAACATCAACGGTAGTAGAAACTTCTTTTACCATGCAAGGACGTGCATCGGAAGGCCGTACCGCGAAAACTTCTTTAG T K Y N G D D K F F I T R E Q V Y G E P M A S K F F D	48 0 486
481 481	V L F H V I R V A R E A L H A V G Q G G H Q G R P R H GTGCTGTTTCATGTGATCCGGGTAGCGAGAGAAGCACCCCGTAGGTCAGGGTGGTCACCAGGGTGGCCACCGCACCAGCACCAGCGACCAGCGACCAGCCAG	560 560
561 561	R Q F A G G T D E F Q G Q F A V G G I T F T F A G H CCGGCAGTTCCCGGTAGCTCACCTTCACCGTCACACCGGACACCGGCCACCACCGGCCACCACCGCCGACACCGCCG	649 649
641 641	A E F V A V H I A I Q F H Q N R H H A G E Q F F A F G CTGAATTTGTGGCCTTCACATCGCCATCCAGTTCCACCAGAATCGGCACCACGCCGGTGAACAGTTCTTCGCCTTTGGA GACTTAAACACGGCAACTGTCAAGAAGGTAGTCTTTGCAGGGAACCT	720 720

GACTTAAACACCGGCAAGTGTACCGGTAGGTCCAGGTGGTGCTTTTAGCCGGCACTTGTCAAGAAGCGGAAACCT S F K H G N V D G D L E V L I P V V G T F L E E G K S

H H M G T CACCATATGGGTACC GTGGTATACCCATGG V M H T G Figure 3 (continued)

	Neuraminidase (-) sequence map (reverse complement	<u>)</u>
Top	: translation of minus strand	
Bot	tom: translation of plus strand (wt protein)	
1 2	ELSRGSSSLVDGEEGQLGAVGPGPAHGV GAGCYCTCTAGAGGATCCTCATCACTTGTCGATGGTGAAGGSCAGCTCGGCCCGTCGGGCCAGGACCAGCTCACGGTGT CTCGAGAGATCTCCTAGGAGTAGTGAACAGCTACCACTTCCCGTCGAGCCGCGCCCGGTCCTGGTGAGCCACA LERSSG KDITFPLEAGDPWSWSVTD	80 80
81 81	A V H A A K A D A A A G P D G A 1. F G P A S D Q L CGCTGTTCACCCCGCAAAAGCTGATGCTGCCGCCGCTGGTCCAGATGGTGCTCTCTTTGGGCCTCCTGATCAGGTCC GCGACAAGTGCGCCTTTTCGACTACGACGACGGCGACCAGGTCTACCACGAGAGAAACCCGGACGAGAACCAGACGACGACGAC	160 160
161 161	H P K A G P D A V Q A C Q L G V L H E A A G V A G P V ACCCAAAAGCAGGCCGGATCAGTCCAGGCCGGTCAGCTCGGGTGCTGCACGAAGCTGCCGGATCAGTCAG	240 240
241 241	G D G H D V L L H A E A A V G P G P A V G V P D H L E GGTGATGGCCACGATGTCCTGCTTCACGCTGAAGCTGCTGTGTGGTCCCGGTCCAGCCGTTGGGGTCCCAGATCATCTCGA CCACTACCGGTGCTACAGGACGAASTGCGACTTCGACGACACCCAGGGCCAAGCCCCAGGGTCTAGTAGAGCT T I A V I D Q K V S F S S D T G T W G N P D W I M E F	320 320
321 321	A G S A V G A L G P A D P H A V A V L E A E A L H A AGCCGGATCTGCTGTTGGTGCTCTTGGTCCGACACCGCGTCGCGTCACCTCACACTCGAAGCCGTTCACGCCGTCGCGCTAGAACCACCACCACCACCACCACCACCACCACCACCACCA	400 400
4 0 1 401	V G A I G A H G S A A A C A I V G P G V V A E H S A A TAGGGGCCATTGGGGGTACTCGCGAACACTCCGCTGCA ATCCGCGGATACCCCGAGCTGCCTGCACACACGGGCTTGCCAACAGCGGCTTGTGAGGCGACGTACCCGAGCACCCCAACAGCGGCTTGTGAGGCGACGT Y A G N P S M P G C S G T G D N P R P N D G F V G S C	480 480
481 481	D V A D L V F Q V L V E G H P G P V A A V P V V P A H GATGTAGCCGATTGTTCCAGGTTGAAGGACACCCAGGGCCGGTTGCTGCCGTGCAGTTGTTCCGGCACACACTACATGGCTAGACCATAAGGGTCCAAGACCAACTTCCTGTGGGTCCCGGCCAACAGGCGCACGGTCAACAGGGCCGTGT I Y G I Q Y E Ł N Q N F S V W P R N S S H W N D R C V	560 560
561 561	A G D L A G V G V A A A F L V V V V G G V Q L H A L CGCAGGTGATCTCGCGGGGGTAGCAGCTGCATCCTCGTAGTGGGGGGGG	640 640
641 641	HHLALFHLENLVGGLPVAGGAVGHHGEAAAACTTGTAGGAGGCCTGCCCGTTGCTGGGGCCGTCATCACGGTGAAGCATGGTGAACGAAGGAAAAGGTAGAAACTTTTAGAAACTTCTCCGGACGGGCAAGGACCCCGGCAGTAGTGCCACTTCGTVVKGKGKGKAAAGGTAAACTTTTAGAACATCTCCCGAACGGGCAACGACCCGGCAGTAGTGCCACTTCGTVVKGKGKGKGKAAGGTAAACTTTTAGAACATCTCCCGAACGGGCAACGACCCGGCAGTAGTGCCACTTCGTVVKGKKGKT	729 729
721 721		800 800
891 801		889 880

Figure 3 (continued)

881 881	A G A G G P G H A L E P A V V G A G S L A H G A A H Q CAGGCBCTGGGGCGACCAGGGGCAGCTCATCAG GTCCGCGACCAGGGCCACCAGGGCCAGCGGCAGCTCATCAG GTCCGCGACCGCCGACCACCAGGGCCAGCCTCGAGAGCTCCCCGACCCTCGAGCGGGTGCCCGTCGAGTAGTC C A S A S W A V S E F R S N Y P S P A E G V P C S M L	960 960
961 961	G P V G A S V L H G A I A V L V V Q Q C S L G Q E E G GGTCCGGTGGGGCTTCTTCCTTCACGGTCCCATGCTCTTCTCTTCACGATGCCAGGAAGAAGGCCCAGGCCACCCCGGAAGAAAGGAAGGAACAAGGAACAGGAAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAACAGGAAGGAAGAA	1949 1949
1041 1041	PAFQVAAADEGCTCAGGGGCTCAGGGCTCCGGGATCACGAACACGTCGCCCTTGCTGCCGATCCGGATCTTGAGGCCCTAGGGCTTAGGCCTAGGCCTAGGCCTAGGCCTAGGCCTAGGCCTAGGCCTAGGCCTAGGCCTAGGCCTAGGCCTAGGCCTAGGCCTAGGCCTACAACRCCGCGAACGACGACGACGACGACGACGACGACGACGACGA	1120 1120
1121 1121	V V L A V H G P A P D G A Q A A V A G Q G H A S H G L TTGTCCTTGCCGGCCAGGGCCAGGGCCACGGCCTTGCCGGCAACGGCCAGGGCCACGGCCAGGGCCACGGCCTTGCCGAACACGAACGA	1200 1200
1201 1201	L G Q G V G V A D A L G L V L A A S L D A V G H P D A CTCGGTCAGGGGTTTGGTTGTTGTGTGTTCTCGGCCTGGTCTGCTCCGGCTCTGGATGCTGTGGAACACCAGATGC GAGCCAGTCCCCAACCACACGACTACGAGACCCAGACCACCACGAC	1280 1280
1281 1281	D H V A D L Q H Q A H G A D H H A D A A D G D D L L TGATCATGTTGCCGATCTGCAGCATCAGCGTCACCATGCAGATGCTGCCGATGGTGATGATCTTCTGG ACTAGTACAACGGCTAGACGTCGTAGTCCCAGTGCCAGGGCTACCACTACTAGAAGACCC I M N G I Q L M L S V T G I V M C I S G I T I I K Q	1360 1360
1361 1361	V G V H G G A A A I S G T TTGGGGTTCATGGTGCGGCGCGCGGATATCGGGTACC 1398 AACCCCAAGTACCALCGGCGCGCGCATAGCCCATGG 1398 N P N M T A R G R Y R T G	
C) M	BD4(-) sequence map (reverse complement)	
Top:	translation of minus strand	
Botte	om: translation of plus strand (wt protein)	
1	E L E S S A Q G Q L L V V L P Q P Y V V L V Q L V V L GAGCTGEAGTCATCAGCTCAGGACACCTCTCGTGGTTCTTCCCCACAGGCCAGTCGTGGTACTTCTTCAGCTTTGTGGTCCTCTCGAGCTCAGTCAG	59 89
81 81	G V H L L P L V H A E D P V A V V A V Ł A D A V Q L CGGGGTGCCCTCCACCCGTGCACGCCACGCCACGCCACG	160 160
161 161	D G V L P L L G Q V L V A E L H D G L G P Q V V Q A Q ATGGGGTACTTCCACGATGGTCATGGCCCTCAGGTCGAGGCCAG TACCCCATGAAGGTGACGAACCAGTCATGAAGAGCAGCCAAGGTCAAAGGTGCAACCAGGAACAAC	240 240
241 241	G L Q Q L A H V P P V G G P G H L G A G V L F Q E L P GEGETTCAGCAGCTCGCCAGTCCGCGGTCCGCGGTCCAGCCACCTCGGCGCGCTCGGGGTACTTTCCAGGAACTTCCCCGAAGTCGTCGACGAGCGAG	320 320

Figure 3(continued)

321 321	Q H G D G H L A A G P V Q K D G G D Q Q L P G V V E ACAGCACGGGGGATGGCCATTGCCGGTGGAACGATGGTGGGAACGACGACGTTCAGGAGGTCCTAGGAGACGACCTTGCAGGACCAGCACCTTGCAGGACCAGCACCTTGCAGGACCAGCACCTTGCAGAGGTCCCCAGGACCTTGCAGGACCTTGCAGAGGTCCCCAGCACCTTGCAGGACCTTGCAGAGGTCCCCAGCACCTTGCAGGACCTTGCAGGACCTTGCAGGACCTTGCAGGACCTTGCAGGACCTTGCAGGACCTTGCAGGACCTTGCAGGACCTTGCAGGACCTGAGACGACCTTGCAGGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCTGAGACGACCAGACACACAC	400 400
401 401	Q G F L D Q V E G A S G G G P L L E G L P P G G A Q G AGGETTTCCTGGACCAGGTTGAAGGGGTTCTGGGGGGGGTCACTTCTTGAAGGCCTTCCGCCTGGGGGGGG	489 489
481 481	L F V V L A A E V Q A G L S P L D L G P G D G V L G E CTCTTTGTTGTACTTGCTGCTGAAGTACAGGCTGGTCTTTCTCCGCTCGATCTGGGTCCAGGGGGGATGGTGCTCGGGTGA GAGAACAACATGAACGACGACGACTTCATGTCCGACCAGAAAGAGGGCGAGCTAGACCCAGGCCCCTAGCACAGGAGCCACT E K N Y K S S F Y L S T K R R E I Q T R P I T D E T F	560 560
561 561	V L P G G A A V V V H L A A P L Q D V G V Q V L F P AGTCCTTCCGGGTGGCGTGCAGTTGTTGTCCATGTCGCTGCCCCTCTTCAGGATGTCGGTGGCAGTGCTGTTCGC TCAGGAAGGCCCACCCCGACGTCAACAACAGGTAGAGCGACGGGAGAAGTCCTACAGCCACCTCCACGAGAAAGGCG D K R T P S C N N D M E S G R K L I D T H L H E K R	640 640
641. 641	L H H L H L G A D F L A F Q E G V L V L L V V E A V L TECACEACETTGECECCATTTCCTCGCTTTCCAGGAAGGTGTCCTCGTACTTCTCGTTGTGCTCGTCCTTTAGGTGCTGGAACCACGGCTAAAGGAGCGAAAGGTCCTTCCACGAGGACCATGAAGAGCAACACGAGCAACACGACGAAGGAAG	720 720
721 721.	G A A E L V D D A A G F L L A A E V A A A Q A P L F GEGGCTGCAGAACTTGTTGATGATGATGCTGCTGCTTGCTGCTGCAGAACTTGTTGATGATGATGATGATGATACTACGGCGACGACGACGACTCTTCAACGACGACGACGACTCCTGAGGCGAGAAAAAAAA	500 500
801 801	L L H Q A V F L A G H A Q C L A A G S G V A D A H G TETTETTCACCAGGETETTTCCTCGCTGGTCGTCGATGTCTCGCCGAGGCTCCGGCGTCGATGCACACGGTC AGAAGAAGTGGTCCGACAAAAGGAGGGACCACTGCGAGTCACAGAGCGGCTCCGAGGCCGAGCGACTACGTGTGCCAG K K V L S N E E S T V S L T E G C A G A D S I C V T	889 889
881 881	P V Q L A L E G H G L A L G V G L V A H G L P L A V A CUSTOCAGCTEGECTTCTCGGCGTCCCCTCCGGCTTGTTGCACACGGACTCCCGCTTGCTGCCTGC	959 959
961. 961	L H E A A A A L P A A L L G F L D G D A L Q D G H L A CTGCACGAAGCCGCTGCAGCTCTTCCTGCAGCCCTTCTTGCTTTTCTTGATGGGGATGCCCTTCAGGATGGTCACCTTGCGACGTGCTTCTCGGGGAAGCACAAAGAACTACCCCTACGGGAAGTCCTACCAGTGGAACGQ V F G S C S K R C G K K T K K I P I G K L I T V K G	1.940 1040
1041 1041	L G L P H F P E V H V V D A L I L F Q Q Q V G A G E CCTTGGGCTCCGCACTTCCGGAAGTCGCTGAAGGCCCTCATCCTCTTTCAGCAGCAGGTGGGTG	1120 1120
1121 1121	V A Q A P A F L Q F G A R A G G H E H V F L A L G P G TTGCTCAGGCCCCTGCTTTCCTGCAGTTCGGAGCTGGGGGGCATGAACACGTCTTTCTT	1200 1200
1201 1201	PQVPVAVVALVLQVA6QE6HAAVLVPA CCGCAGGTTCCAGTTGTTGTTCTGTTTGGTCTGGTGCGGGCGG	1280 1289

Figure 3 (continued)

1281 1251	L D A P L G Q H G E V E V L G L Q A C L A V L V Q V TCTTSATGCCCGGCTTGAGCACGGTGAAGTCGAGGTCTTCAGGCTTGATGCCCCGCGTTCTTGTGCAGGTAGAGACTACGGGGGAACCTGTCGTGCCACTTCAGCTTCAGGAGCCCGAAGTCCBAACAAGAGCGCAAGAACACGTCCATC K I G R K S L V T F D F D E P K L S T E G N K H L Y	1360 1360
1361 1361	V 6 Q A A L A S E L Q A L G A D E V H V E S A G G L A TTGGCCAGGCTCTTTCTTCTTCTCAACTTCAAGCCCGCGCTCTACTTCATCTCCCGCCGCCGTCTTGCCAACCGGTCGAACAGAACGAAACGAAACGAAACGAACG	1440 1440
144 1 1441	E Q P L L H H P L P A A G H A L P T L G S G G G A E L GAACAGCCCTTCTCACCACCCCCCTCCCAGCCCCCCCCCC	1520 1520
1521 1521	G A G D R F L Q Q G V A L A A P L D H H L F L V L A EGECCCTGGCGATAGGTTCCTGCAGGAGGGGGTTGCACTGCTCGCTGCTGCTGCTGATCATCATCATCTGTCCTCGTCCTCGCCC CCCGCGACCGCTATCCAACGACGACGGTCGTCCCCCAACGTGAGCGACCAGGAGCAACTAGTAGTAGAAAAGGAGCAGGAGGGG	1600 1600
1601 1661	H S F Q F H G H I F L P Q V V G G V G H Q P L A A G H ACTUTTCCAGTTCCATGGCCACATCTTCCTTCCGCAGGTCGTTGSGGGGGTCACCAGCCGCTCGCTGGTCACTGGTCACTGGTCACAGGTCAAGGTCAAGGTCAAGGTACAGGTAGAAGGAAG	1680 1680
1681 1681	G R G G S A V A Q A Q A F Q A G G A H E G K L G T GGTAGGGGGGGCTCGCCCAGGCTCAGGCTTCCAGGCCGGTGCTGCCAAACCTTSGTACC 1755 CCATCCCCGCCGAGGCGCCAGGGGTCGGAAAGGTCCGAAAGGTCCGCCACCAGGGTACCACCGTTCGAACCATGG 1755 T P A A G R D G L S L S E L G T T G M T A L K T G	

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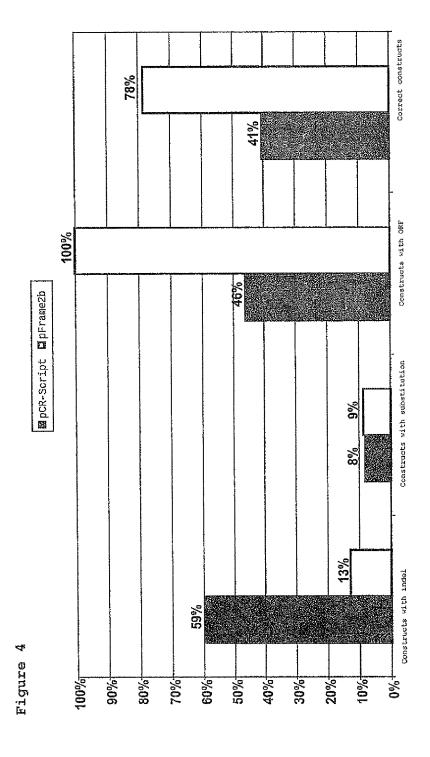


Figure 5

Partial sequence of the pFroup1 vector

Figure 5 (continued)

K TELA TO WELLE E YER DESEGENTLY DE ALLAS VER BAAAACAGCATTCCAGGTATTAGAAGAATATCCTGATTCAGGTGAAAATATTGTTGATGCGCTGGCAGTGTTCCTGCGCCTTTTTGTTGTTGAAGGTCCATAATCTTCTTATAAGACTAAGGTCCACTTTTATAACAACTACGCGACCGTCACAAGGACGCGG

GGTTGCATTCGATTCCTGTTTGTAATTGTCCTTTTAACAGCGATCGCGTATTTCGTCTCGCTCAGGCGCAATCACGAATGCCAACGTAAGCTAAGGACAAACATTAACAGGAAAATTGTCGCTAGCGCATAAAGCAGAGCGAGTCCGCGTTAGTGCTTAC

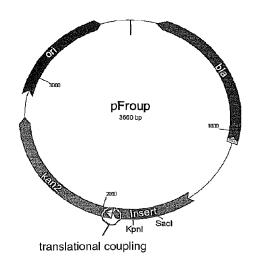
K CALLA PACE SA PADA SA VALATA HAGADA FASALLADAN ALLATA FADA ESGAM TAMACTITIGCCATTCTCACCGGATTCAGTCGTCACTCATGGTGATTCTCACCTGATAACCTTATTTTTGACGAGGGGAATTTGAAAACCGGTAAGAGTGGCCCTAAGTCAGCAGTGAGTACCACTAAAGAGTGAACTATTGGAATAAAAACTGCTCCCCT

LONGO GOLO VO GOROVO GOLO BOLO A GOLO WINNESSE AATTAATAGGTGTATTGATGTTGGACGAGTCGGAATCGCAGACCGATACCAGGATCTTGCCATCCTATGGAACTGCCTCTTAATTATCCAACATAACTACAACCTGCTCAGCCTTAGCGTCTGGCTATGGTCCTAGAACGGTAGGATACCTTGACGGAG

G F F S P S F 0 K R F F 00 K Y G F D N 12 D M N K E 0 F G TGAGTTTTCTCCTTCATTACAGAAACGGCTTTTTCAAAAATAGGTATTGATAATCCTGATATGAATAAATTGCAGTTCCACTTCAAAAAGAGGGAAGTAATGTCTTTGCCGAAAAAGTTTTTATACCATAACTATTAGGACTATACTTATTTAACGTCAA

HILDING DIE RESERVE SELVNWL*HWQSITLT*
TCATTTGATGCTCGATGAGTTTTCTAATCAGAATTGGTTAATTGGTTGTAACACTGGCAGAGCATTACGCTGACTTGAC
AGTAAACTACGAGCTACTCAAAAAAGATTAGTCTTAACCAATTAACCAACATTGTGACCGTCTCGTAATGCGACTGAACTG

Figure 6



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Figure 7

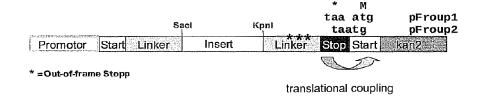
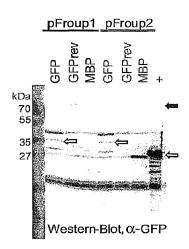


Figure 8



METHOD FOR DETERMINING FRAMESHIFT MUTATIONS IN CODING NUCLEIC ACIDS

SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted via. EFS-Web and is hereby incorporated by reference in its entirety. Said ASCII copy, created on Jul. 28, 2010, is named WEICKM86.txt, and is 76,815 bytes in size.

The present invention relates to a method for identifying frameshift mutations in coding nucleic acid sequences.

Genetic information is organised in coding nucleic acids in the form of base triplets or codons. However, errors frequently occur in gene synthesis in the form of various kinds of mutations. The majority of the mutations are single-base deletions and insertions (approx. 90%). In contrast with most substitutions, virtually all insertions and deletions give rise to what are known as frameshifts. If a base is lost from a gene in the course of a mutation (deletion) or if a base is added (insertion), this changes the reading frame of the following base triplets. A frameshift mutation has a substantially greater biological impact than a substitution. From the insertion/ 25 deletion onwards, other amino acids are encoded and translation is usually terminated due to out-of-frame stop codons.

As the length of a synthesised gene increases, the probability of a mutation occurring likewise greatly increases. The majority of unwanted mutations occur due to the absence or 30 the insertion of individual bases into oligonucleotides. The probability of a base being substituted during the synthesis of genes or gene libraries amounts to approximately 0.1-0.2% per position, while the probability of an insertion/deletion is 1.0-1.5% per position. For example, Kong et al., Nucleic 35 Acids Res. (2007), 35:e61 observed a deletion rate of 1.48% per position and a substitution rate of 0.3% per position in chip-based gene synthesis. Around half of the substitutions were in turn due to PCR errors.

The reading frameshift which occurs in the event of insertions and deletions may be exploited in order, by means of a reporter vector, to select those nucleic acid fragments which have an intact reading frame. European patent application EP 0 872 560 accordingly discloses a method for identifying frameshift mutations, in which homologous recombination is used to produce a construct which contains a promoter, a gene to be investigated and a reporter gene in the same reading frame as the gene to be investigated. Upon expression of the reading frame, a fusion protein is obtained which contains the reporter gene function. The latter may be detected by means 50 of phenotypic characteristics.

One drawback, however, is that, for the procedure described in EP 0 872 560, the gene to be investigated must not comprise any internal stop codons within the open reading frame (ORF). However, many synthesised genes do comprise 55 such internal stop codons.

In the light of the above problem, the object of the present invention was to provide a method for identifying frameshift mutations which is suitable for any coding nucleic acids and in particular also for nucleic acids which comprise internal 60 stop codons.

According to the invention, said object is achieved by a method for identifying frameshift mutations in coding target nucleic acids which comprises the steps:

(i) providing a host cell comprising a double-stranded nucleic 65 acid, which comprises a coding target nucleic acid and a coding opposite strand nucleic acid complementary

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thereto, in which the opposite strand nucleic acid is present in operative linkage with a reporter gene in 3'-position;

- (ii) effecting expression of the opposite strand nucleic acid; and
- (iii) identifying whether expression of the reporter gene occurs in the host cell,

in which expression of the reporter gene indicates that the target nucleic acid does not comprise a frameshift mutation.

According to the invention, the phrase "does not comprise a frameshift mutation" should be taken to mean that the target nucleic acid either comprises absolutely no frameshift mutation or that a plurality of mutations are present in the target nucleic acid, which, viewed in isolation, would result in a reading frameshift, but cancel each other out again. For example, a combination of insertions and deletions may be present in the target nucleic acid which however cancel each other out again, such that the reading frame is not modified and the reporter gene may still be correctly read.

The inventors have found that, instead of the coding target nucleic acid, a coding opposite strand nucleic acid complementary thereto may be used to identify frameshift mutations.

In this connection, the term "coding" should be taken to mean that the opposite strand nucleic acid is such that it enables expression of the 3'-linked reporter gene. However, the opposite strand nucleic acid need not necessarily be defined for this purpose by an open reading frame.

When the method according to the invention is carried out, a host cell is first provided which comprises a double-stranded nucleic acid which contains a coding target nucleic acid and a coding opposite strand nucleic acid complementary thereto. The target nucleic acid preferably comprises a synthetically produced sequence. In the host cell, the opposite strand nucleic acid is in operative linkage with a reporter gene which is located downstream in 3'-position.

If the opposite strand nucleic acid is intact, i.e. comprises no mutations such as in particular insertions or deletions which result in a modification of the reading frame, on expression a fusion polypeptide is obtained which comprises the amino acid sequence coded by the opposite strand nucleic acid and, C-terminally therefrom, the amino acid sequence of the product coded by the reporter gene. If, on the other hand, the opposite strand nucleic acid does comprise a frameshift mutation, i.e. in particular an insertion or deletion which modifies the reading frame, this reading frameshift results in the reporter gene not being located in-frame relative to the coding opposite strand nucleic acid. As a result, the reporter gene is not expressed. The reporter is thus only expressed if the reading frame of the opposite strand is intact.

If the opposite strand nucleic acid does not comprise any frameshift mutations, it may be concluded that the complementary target nucleic acid itself is also intact and does not comprise any frameshift mutations.

The method of the present invention has the advantage that, even if internal stop codons are present in the target nucleic acid, it is possible to identify frameshift mutations by using a reporter gene. When the reading frame in the opposite strand is used, internal stop codons of the target nucleic acid are translated as Leu or Ser. Using a reading frame in the opposite strand furthermore has the advantage that any possible toxicity of the protein coded by the actual target nucleic acid for the host cell is irrelevant, as only the opposite strand is translated.

A coding opposite strand nucleic acid is necessary in order to carry out the method according to the invention. A reading frame in the opposite strand may optionally be obtained by appropriate optimisation of the opposite strand sequence. It may furthermore be preferred for the opposite strand nucleic acid to contain no internal stop codons. The complementary

codons to the three stop codons are relatively rare, so a reading frame in the opposite strand generally contains no stop codons. In a preferred embodiment the opposite strand nucleic acid is optionally optimised such that no internal stop codons are present.

In a preferred embodiment according to the invention, the opposite strand nucleic acid in the host cell in 3'-position is in operative linkage with a reporter gene and in 5'-position is in operative linkage with an expression control sequence.

According to the invention, an expression control sequence is a nucleic acid sequence which controls and regulates transcription and translation. The expression control sequence may be constitutively or regulatably active in the host cell. The phrase "in operative linkage" includes a suitable start signal (for example ATG or AUG) before the nucleic acid sequence to be expressed and the retention of the correct reading frame, so enabling expression of the nucleic acid sequence under the control of the expression control sequence and the production of the product coded by the nucleic acid sequence. If a nucleic acid does not contain a suitable start signal, such a start signal may be inserted before the nucleic acid.

In one embodiment, the provision of a host cell in step (i) of the method according to the invention involves the introduction of an expression vector into a host cell, wherein the 25 expression vector comprises the double-stranded nucleic acid which comprises a coding target nucleic acid and a coding opposite strand nucleic acid complementary thereto.

Expression vectors are known to a person skilled in the art in the field of molecular biology and are described, for 30 example, in Sambrook et al., Molecular Cloning, A Laboratory Manual (1989), Cold Spring Harbor Laboratory Press. For example, a plasmid or a viral vector may be used as expression vector.

In one embodiment, the opposite strand nucleic acid may 35 be present in the expression vector in operative linkage with a reporter gene in 3'-position. Alternatively, in another embodiment, the expression vector may be introduced into a host cell together with a reporter vector which comprises the reporter gene. In a further embodiment, the expression vector 40 may also be introduced into a host cell which already contains a reporter gene.

The opposite strand nucleic acid in the expression vector is preferably in operative linkage with an expression control sequence in 5'-position. In a particularly preferred embodiment, the opposite strand nucleic acid in the expression vector is in operative linkage in 3'-position with a reporter gene and in 5'-position with an expression control sequence.

An expression vector according to the invention may be produced in any desired manner, for example by culturing in 50 host cells, preferably in bacterial cells such as for instance *E. coli* cells. The expression vector conveniently contains elements which enable replication and selection in the host cell. Alternatively, an expression vector may also be produced in vitro by amplification in sufficient quantity, for example by 55 polymerase chain reaction (PCR), ligase chain reaction (LCR) or rolling circle amplification.

Prokaryotic or eukaryotic cells or microorganisms may be used as host cells. The host cells may be wild-type variants or mutated or genetically manipulated host cells may be used. 60

According to the invention, a gene which codes for a detectable gene product may be used as a reporter gene. For example, the reporter gene may comprise an antibiotic resistance gene, preferably kanamycin resistance. In another embodiment, a reporter gene may be used which codes for a 65 fluorescent protein such as for instance GFP. It is furthermore possible to use a reporter gene which codes for an enzyme

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which catalyses a colorimetrically measurable reaction (for example β -galactosidase). In a further embodiment, the reporter gene may code for a gene product which interacts with a nucleic acid sequence present in the host cell or with a gene product expressed in the host cell, whereby a measurable change in metabolic activity comes about.

The expression vector and optionally reporter vector is/are introduced into the host cell in accordance with methods known in the prior art, for example by (co)transfection, (co) transformation or (co)infection of cells. The method is preferably selected such that the double-stranded nucleic acid is introduced into the host cell in such a way that, in the host cell, the opposite strand sequence is in operative linkage with the reporter gene.

In eukaryotic host cells, transfection or cotransfection may, for example, proceed by calcium phosphate coprecipitation, lipofection, electroporation, particle bombardment, by using bacterial proteins or by viral infection via retroviruses, adenoviruses etc.

In a preferred embodiment of the invention, in step (i) of the method a vector is provided which, in addition to the reporter gene in operative linkage with the opposite strand nucleic acid, comprises at least one selection marker gene. The at least one selection marker gene is selected from any desired genes which code for a detectable gene product, wherein the gene product preferably differs from the gene product of the reporter gene. For example, the selection marker gene may comprise an antibiotic resistance gene such as for instance β -lactamase for ampicillin resistance. Alternatively, a gene which codes for a fluorescent protein such as for instance GP may be used as a selection marker gene.

Selection may proceed independently of the reporter gene by means of the selection marker gene. For example, after selection for the first reporter gene (for example for kanamycin resistance), a construct may be further up-amplified by selection by means of the further selection marker gene (for example for ampicillin resistance). It is furthermore possible to further amplify constructs which contain a frameshift mutation (for example an intentional frameshift mutation) by selection via the second selection marker gene.

Finally, the second selection marker gene may assist in disguising the existence of the reporter gene in connection with a selection which has taken place, in order to protect the method from imitation.

Step (ii) of the method according to the invention involves effecting expression of the opposite strand nucleic acid in the host cell. To this end, the host cell is cultured under suitable conditions which permit expression of the opposite strand nucleic acid.

Step (iii) involves identifying whether expression of the reporter gene occurs in the host cell. The identification is based on the detection of a gene product coded by the reporter vector. Depending on the reporter gene used, the identification may in principle comprise the identification of any phenotypically recognisable effects, for example morphological changes, changes to growth behaviour, etc. If, for example, the reporter gene codes for a fluorescent protein such as GFP, the presence and/or the intensity of luminescence may for example be identified by fluorescence cytometry or imaging assays. If an antibiotic resistance gene is used as reporter gene, the identification may be made on the basis of the growth of host cells in the presence of antibiotics.

In one embodiment of the invention, a target nucleic acid may be assembled from two, three or more subfragments. For example, in the case of a 3 kB construct, three subfragments may first be produced from oligonucleotides and these subfragments then fused to form a 3 kB fragment, for example by

fusion PCR. A construct fused in this manner from two or more subfragments may then be ligated directly into an expression vector according to the invention.

Since the method according to the invention is capable of identifying frameshift mutations, but not mutations where the reading frame is maintained, in the event that the target nucleic acid is synthesised by PCR the amplification primer is preferably added at the latest possible time in order to keep the number of amplification steps as small as possible. The greater the number of amplifiable full-length molecules, the fewer the mutations arising due to PCR which will be present in the PCR product.

Ofloxacin, for example, may be used after transformation in order to reduce substitutions still further. Ofloxacin is a gyrase inhibitor which inhibits DNA replication but, over the short term, does not kill host cells such as *E. coli* cells. During this period, a host cell has the opportunity to cleave heterodimers via an internal repair mechanism. Ofloxacin may preferably be present in the medium after electroporation or heat-shock transformation (0.5-1 h at 37° C.). The cells are then centrifuged off, the medium together with the ofloxacin removed and the cells are plated out.

The above-stated method is based on the fact that only in the absence of frameshift mutations does translation of the 25 opposite strand give rise to a fusion protein which comprises the reporter. Under certain circumstances, due to its size, such a fusion protein has an elevated molecular weight and may be present in the cell in the form of denatured aggregates. Even if the correct reading frame is inserted, the expression and functionality of the reporter may thus be affected or even inhibited by the properties of the polypeptide formed. This may lead to a reduction in the efficiency of the method, in particular in the case of relatively large coding target nucleic acids and corresponding opposite strand nucleic acids.

The previously disclosed procedure was further developed by the inventors in the light of this problematic issue. It has surprisingly been found that the strategy of translational coupling known in the field may be used in the above method of operative linkage of the coding opposite strand nucleic acid with a reporter gene in 3'-position.

The phenomenon of conjugated translation or translational coupling is a control mechanism in which the translation of an upstream gene regulates the translation of a downstream 45 gene. One theory to explain this phenomenon assumes that the ribosome from the translated upstream gene is passed on to the downstream gene via a translational coupling signal which acts as a weak ribosome-binding site. The ribosome is not rebound here, but instead, once translation of the 50 upstream gene is complete, the ribosome can scan the sequence and initiate translation of the downstream gene. Translational coupling between two cistrons is thus mediated via the same ribosome. The ribosome terminates translation at a stop codon in the upstream sequence and thereupon scans 55 the downstream sequence, beginning the new translation at a start codon in the vicinity of this stop codon. This scanning operation by the ribosome proceeds in both directions, such that continuation of synthesis may be initiated at a start codon which overlaps with the stop codon of the preceding coding 60

Translational coupling was described for the first time by Oppenheim 1980 for the tryptophan operon in *E. coli* (D. S. Oppenheim and C. Yanowski (1980), Genetics 95:789-795). The effect is based on the fact that translation restarts after a 65 stop codon where there is a start codon following directly thereafter. This gives rise to two separate polypeptide chains.

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A prerequisite for the synthesis of the second polypeptide is that the first reading frame is correctly read up to the stop coden

WO 2008/077881 describes a method for selecting genes from a gene library for improved expression efficiency, improved expression being quantified by means of a reporter gene which is synthesised by translational coupling with the corresponding gene. Genes with an open reading frame are simultaneously identified by this method. E.g. resistance genes or GFP are mentioned as reporters. It is additionally noted that the start codon of the reporter gene may both overlap with the stop codon of the open reading frame and be at a distance of up to 500 nucleotides.

WO 2008/051619 describes a method for screening DNA libraries for identifying DNA fragments with an open reading frame which comprise neither internal ribosomal binding sites nor internal stop codons. The sequences are selected by using reporter proteins which are not synthesised by covalent fusion with the corresponding reading frame, but instead by translational coupling, in order to prevent misfolding or malfunction of a corresponding fusion protein. The document additionally discloses two vector variants which either enable positive selection of open reading frames without stop codons via resistance markers, or effect negative selection of open reading frames with internal ribosomal binding sites by expression of a toxin.

Ohashi-Kunihiro et al. (Biotechniques (2007) 43(6):741-2, 754) likewise describe the selection of DNA fragments with an open reading frame by translational coupling with a resistance marker which is only expressed in the absence of internal stop codons. Further, the optimal distance between the open reading frame and the resistance marker is determined.

It has now surprisingly been found in the present invention
that the strategy of translational coupling may be used in the
previously disclosed method for selecting coding nucleic acid
constructs for the absence of frameshift mutations. However,
in contrast with the method described in the prior art, in the
method according to the invention the open reading frame of
the opposite strand of the coding target nucleic acid is
selected by translational coupling.

The invention therefore provides a method for identifying frameshift mutations in coding target nucleic acids which comprises the steps:

- 5 (i) providing a host cell comprising a double-stranded nucleic acid, which comprises a coding target nucleic acid and a coding opposite strand nucleic acid complementary thereto, in which the opposite strand nucleic acid is linked via a linker with a reporter gene in 3'-position;
- (ii) effecting expression of the opposite strand nucleic acid;
 and
- (iii) identifying whether expression of the reporter gene occurs in the host cell,
- in which expression of the reporter gene indicates that the target nucleic acid does not comprise a frameshift mutation, characterised in that the linker comprises a translational coupler sequence which comprises a stop codon in frame to the reading frame of the opposite strand nucleic acid and a start codon, wherein the reporter gene is located in frame to the start codon.

If the opposite strand nucleic acid (and thus also the complementary target nucleic acid) does not comprise a frameshift mutation, the opposite strand may be correctly read up to the stop codon of the translational coupler sequence in the linker bound in 3'-position. Translation then restarts at the subsequent start codon of the translational coupler sequence. Since the start codon in the linker used according to

the invention is in frame to a subsequent reporter gene, the reporter gene is in this case also translated.

By using translational coupling in the method of the present invention, the expression product of the opposite strand nucleic acid to be checked and of the reporter gene are 5 accordingly obtained as separate polypeptide chains.

With the assistance of the method according to the invention, even if internal stop codons are present in a target nucleic acid, frameshift mutations may successfully be identified by means of a reporter gene by using the opposite strand nucleic 10 acid. When the reading frame in the opposite strand is used, internal stop codons of the target nucleic acid are translated as Leu or Ser. Using a reading frame in the opposite strand furthermore has the advantage that any possible toxicity of the protein coded by the actual target nucleic acid for the host 15 vector is provided which, in addition to the reporter gene cell is irrelevant, as only the opposite strand is translated.

According to the invention, a translational coupler sequence comprises a stop codon, which is arranged in frame to the opposite strand nucleic acid, and a subsequent start codon, which is arranged in frame to the reporter gene. The 20 distance between the stop codon and start codon is selected such that translational coupling is enabled. The distance preferably amounts to no more than 10 base pairs.

In one embodiment of the present invention, the start codon and stop codon of the translational coupler sequence follow 25 on immediately from one another. The translational coupler sequence may, for example, comprise two separate codons, for example TAA ATG. In another embodiment, the start codon and stop codon of the translational coupler sequence overlap with one another. One example of an overlap of the 30 stop codon with the start codon is the translational coupler sequence TAATG.

The coupling of translation at the genetic level according to the invention with simultaneous decoupling of the resultant polypeptide chains results in a significant improvement to the 35 method for identifying correct reading frames. The system remains independent of the folding of the polypeptide chain formed by the opposite strand nucleic acid.

In a preferred embodiment of the invention, the linker contains, in the reading frames shifted by +1 and -1, further 40 stop codons located upstream of the stop codon of the translational coupler sequence. These stop codons ensure that translation of the shifted reading frames is terminated before the stop codon of the translational coupler sequence is reached. The distance of the further stop codons from the start 45 codon of the translational coupler sequence is here preferably selected such that no translational coupling occurs. A distance of at least 30 base pairs, preferably of at least 50 base pairs, has proved to be particularly suitable.

In a preferred embodiment, the opposite strand nucleic acid 50 in the host cell in 3'-position is linked via a linker with a reporter gene and in 5'-position is in operative linkage with an expression control sequence.

The procedure in the method according to the invention for identifying frameshift mutations moreover corresponds to the 55 °C. previously described method.

In one embodiment, the provision of a host cell in step (i) of the method involves the introduction of an expression vector into a host cell, wherein the expression vector comprises the double-stranded nucleic acid which comprises a coding target 60 nucleic acid and a coding opposite strand nucleic acid complementary thereto. In the expression vector, the opposite strand nucleic acid is preferably joined with a linker in 3'-position which comprises the translational coupler sequence.

The opposite strand nucleic acid in the expression vector is 65 preferably in operative linkage with an expression control sequence in 5'-position. In a particularly preferred embodi-

ment, the opposite strand nucleic acid in the expression vector in 3'-position is linked via a linker with a reporter gene and in 5'-position is in operative linkage with an expression control

The expression vector and optionally reporter vector is/are introduced into the host cell in accordance with methods known in the prior art, for example by (co)transfection, (co) transformation or (co)infection of cells. The method is preferably selected such that the double-stranded nucleic acid is introduced into the host cell in such a way that, in the host cell, the opposite strand sequence is linked with the reporter gene via the linker which comprises a translational coupler sequence.

In a preferred embodiment, in step (i) of the method a linked via a linker with the opposite strand nucleic acid, comprises at least one selection marker gene.

FIGURES

FIG. 1 shows the plasmid map of an expression vector according to the invention. Kanamycin resistance is present as the reporter gene downstream of the opposite strand nucleic acid MCS. FIG. 1 discloses the amino acid sequence as SEQ ID NO: 17 and the nucleic acid sequence as SEQ ID NO: 16.

FIG. 2 Streaked plates of colonies with kanamycin selection vectors (pFrame2A3b and pFrameT1) with different KpnI/SacI insertions. The insertions on plate A) and B) comprise, on the one hand, beta-lactamase including promoter (A and B, top), which is cloned in reverse orientation to the kanamycin resistance gene and imparts amp resistance but does not permit kana resistance, since the opposite strand of promoter+beta-lactamase does not contain an open reading frame, and, on the other hand, eGFP (C and D), which is present here cloned in identical orientation and in-frame to the kanamycin resistance gene. Here, the continuous reading frame of the plus strand of eGFP enables expression of kanamycin resistance, while the absence of the ampicillin cassette naturally does not permit any growth on ampicillin. These data demonstrate that a selection for sequences with open reading frames is possible with the assistance of the method according to the invention.

In contrast, three colonies are streaked on plates C) and D), which colonies contain three different insertions in negative orientation in the pFrame2A3b kanamycin selection vector, all three also having an open reading frame in the minus strand. The insertions comprise the genes for eGFP (735 bp), influenza neuraminidase (1398 bp) and human CpG-binding protein MBD4 (1755 bp).

All three constructs are thus capable of growing on kanamycin. It is, however, also clear in individual cases, namely in particular MBD4(-) and to a lesser extent also neuraminidase (-) and eGFP(-), that growth is stabler at 30° C. instead of 37°

These three fusion proteins from minus strand translation plus neomycin phosphotransferase are thus capable of imparting phenotypic and selectable kanamycin resistance.

FIG. 3 shows the sequences of the three insertions from FIG. 2 with annotated plus and minus translation. FIG. 3A discloses SEQ ID NOS 2, 1, 3 and 4, respectively, in order of appearance. FIG. 3B discloses SEQ ID NOS 6, 5, 7, 18 and 8, respectively, in order of appearance. FIG. 3C discloses SEQ ID NOS 10, 9, 11 and 12, respectively, in order of appearance.

FIG. 4 illustrates a comparison of the selection of coding nucleic acid constructs for the absence of frameshift mutations with the assistance of a conventional cloning vector and

a cloning vector according to the invention. The grey bars show the result of selection using the conventional pCR Script cloning vector, while the white bars show the result of selection using the pFr2b selection vector according to the invention

FIG. 5 shows a partial sequence of a pFroup1 vector for use in the method of the invention. The opposite strand nucleic acid (insert) was linked via a linker with a kanamycin resistance gene (kan2). The linker shown in each case contains a plurality of stop codons in the reading frames shifted by +1 and by -1 relative to the reading frame of the opposite strand sequence. The linker furthermore contains a translational coupler sequence (TAA ATG), the stop codon of which is in frame to the reading frame of the opposite strand nucleic acid, and the start codon of which is in frame to the kanamycin resistance gene. FIG. 5 discloses SEQ ID NOS 14, 13, 15 and 19-21, respectively, in order of appearance.

FIG. **6** shows the structure of the pFroup1 and pFroup2 vectors according to the invention. Both are designed for use in *E. coli*. The plasmid imparts ampicillin resistance to the ²⁰ host cell, independently of an insert, so ensuring straightforward multiplication. The insert to be investigated is cloned into the vector via the KpnI and SacI restriction sites. Expression is under the control of a constitutive promoter.

FIG. 7 shows the coupling of the insert and kan2. A linker ²⁵ is fused N-terminally to the insert, the linker bearing stops in the +1 and –1 reading frame (out-of-frame stops). In the case of deletions and insertions which shift the reading frame, translation is terminated at this point. Only if the insert is correctly translated is the resistance marker synthesised via ³⁰ translational coupling, which in this case results in kanamycin resistance of the host cell.

FIG. 8 shows the result of an immunoblot analysis of the expression product of a nucleic acid sequence coding for GFP which was inserted into the plasmids pFroup1 and 2. The GFP 35 synthesised in the transformed cells was investigated by immunoblot in order to establish that translational coupling results in the formation not of a large fusion protein, but instead of two separate polypeptide chains. The controls used were cells which had been transformed with GFPrev, the 40 opposite strand nucleic acid of GFP, which codes for a nonsense polypeptide, or MBP. Free GFP has a size of 40 kDa (white arrow), the GFP-Kan2 fusion protein would be 70 kDa in size (black arrow). GFP from a comparison vector is applied as a positive control (+). Since no linker is present 45 N-terminally here, it is only approx. 30 kDa in size. Translational coupling is functional both in pFroup1 and in pFroup2. Accordingly, no detectable Kan2 fusion protein is formed.

The following exemplary embodiments are intended to provide further illustration of the invention.

EXAMPLE 1

Selection for Absence of Frameshift Mutations

A construct produced entirely from oligonucleotides with a length of 684 bp was cloned into the conventional pCR Script cloning vector and into the pFr2b selection vector according to the invention via the restriction enzymes KpnI/SacI. The sequence of the construct was designed by alternative codon 60 selection such that, in addition to the biologically relevant reading frame (with a terminal stop codon), it comprises a second open reading frame in reverse direction. This does not, however, have any effect on the biologically relevant protein coded on the sense strand.

Ligation of the construct (insert) into the pFr2b vector gives rise to an open reading frame within the vector which is

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composed of start+reverse insert+selection gene (kana). Only if the insert's reading frame is intact can kana be correctly translated and enable growth of the cell/colony.

The ligations were transformed into $E.\ coli$ and plated out onto selection plates comprising 50 µg/ml of kanamycin. Plasmid DNA was isolated from individual grown colonies, sequenced and analysed in accordance with the following criteria listed in Table 1.

TABLE 1

	Construct	Vector
	0800349 pCR-Script	0800349 pFr2b
Orientation	Reverse	Reverse
Size	684 bp	684 bp
Peer group	37	23
Total insertions	12	0
Insertions per construct	0.32	0.00
Insertions per kb	0.47	0.00
Total deletions	20	4
Deletions per construct	0.54	0.17
Deletions per kb	0.79	0.25
Constructs with indel	22	3
Constructs with indel (%)	59%	13%
Constructs with in-frame indel	2	3
Constructs with in-frame indel (%)	5%	13%
Total transitions	2	2
Transitions per construct	0.05	0.09
Transitions per kb	0.08	0.13
Total transversions	1	0
Transversions per construct	0.03	0.00
Transversions per kb	0.04	0.00
Constructs with substitution	3	2
Constructs with substitution (%)	8%	9%
Constructs with ORF	17	23
Constructs with ORF (%)	46%	100%
Constructs without indels	15	20
Constructs without indels (%)	41%	87%
Correct constructs	15	18
Correct constructs (%)	41%	78%

	Inde	ls	
	pCR script	pFr2b	
>3 bp del (size)	3 (8 bp, 591 bp, 456 bp)	0	
3 bp del	0	2	
2 bp del	1	1	
1 bp del	15	1	
1 bp ins	12	0	
2 bp ins	0	0	
3 bp ins	0	0	
>3 bp ins	0	0	

The terms and abbreviations used in Table 1 above have the following meanings:

Construct Name of the construct. The gene was in each case cloned into the vectors via KpnI/SacI.

Vector pCR-Script: conventional cloning vector; pFr2b: selection vector according to the invention for open reading frames

Orientation Orientation in which cloning into the vector was performed. Only of relevance to pFr2b. Cloning was therefore performed here such that the reverse reading frame of the gene is present in fused form with the selection cassette. Size Length of the gene in bp.

Peer group Number of sequenced clones.

Total insertions Total number of insertions found in the random sample.

Insertions per construct Calculated insertions per construct. Insertions per kb Calculated insertions per kilobase.

Total deletions Total number of deletions found in the random sample.

Deletions per construct Calculated deletions per construct. Deletions per kb Calculated deletions per kilobase.

Constructs with indel Total number of constructs found in the random sample with at least one insertion or deletion. In the case of pFr2b, this is not identical to the sum of all insertions deletions, since in this case there is one construct with two deletions, for example.

Constructs with indel (%) Percentage of constructs in the 10 random sample with at least one insertion or deletion. Distinctly fewer constructs with indels are found in the case of pFr2b (13% vs. 59%). See FIG. 4.

Constructs with in-frame indel Total number of constructs found in the random sample with at least one insertion or deletion which, however, does not interrupt the reverse open reading frame. It should be noted that in the case of pFr2b the three constructs with an indel still nevertheless comprise an intact reverse open reading frame. The reason for this is that two constructs have a 3 bp deletion, one 20 construct has a 1 bp deletion and shortly thereafter also a 2 bp deletion; these do not interrupt the reading frame since the deletions are divisible by three, as is mentioned in the general part of the description.

Constructs with in-frame indel (%) Percentage of constructs 25 found in the random sample with at least one insertion or deletion which, however, does not interrupt the reverse open reading frame.

Total transitions Total number of transitions found in the random sample (purine <-> purine or pyrimidine <-> 30 pyrimidine).

The fact that transitions are more frequently observed than transversions indicates that the substitutions were not introduced by oligonucleotide errors, but instead by PCR errors.

Transitions per construct Calculated transitions per construct. Transitions per kb Calculated transitions per kilobase.

Total transversions Total number of transversions found in the random sample (purine <-> pyrimidine).

Transversions per construct Calculated transversions per con-

Transversions per kb Calculated transversions per kilobase. Constructs with substitution Total number of transitions+ transversions (=substitutions) found in the random sample.

Constructs with substitution (%) Percentage of constructs in 45 the random sample with at least one substitution. No selection is made by pFr2b with regard to substitutions. See FIG.

Constructs with ORF Total number of constructs found in the random sample with a reverse open reading frame.

Constructs with ORF (%) Percentage of constructs in the random sample with a reverse open reading frame. It should be noted that this value is 100% in the case of pFr2b. See FIG. 2.

Constructs without indels Total number of constructs found in 55 the random sample without an insertion or deletion.

Constructs without indels (%) Percentage of constructs in the random sample without an insertion or deletion. It should be noted that this value is 87% in the case of pFr2b, i.e. more than twice as high as without corresponding selection.

Correct constructs Total number of constructs found in the random sample which have neither an insertion or deletion, nor a substitution, which are thus 100% correct.

Correct constructs (%) Percentage of constructs in the random sample which have neither an insertion or deletion, nor a substitution, which are thus 100% correct. It should

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be noted that this value is 87% in the case of pFr2b, i.e. almost twice as high as without corresponding selection. See FIG. 4.

>3 del (size) Total number of deletions of >3 bp found in the random sample and their size.

3 del Total number of deletions of 3 bp found in the random sample.

2 del Total number of deletions of 2 bp found in the random sample.

1 del Total number of deletions of 1 bp found in the random sample.

1 ins Total number of insertions of 1 bp found in the random sample.

2 ins Total number of insertions of 2 bp found in the random sample.

3 ins Total number of insertions of 3 bp found in the random sample.

>3 ins Total number of insertions of >3 bp found in the random sample.

Result:

In the example given, cloning into the pFr2b selection vector leads to 100% elimination of those inserts whose reading frame has been destroyed by insertions or deletions. However, no selection occurs for the far rarer substitution mutations. Insertions/deletions which leave the reading frame intact are likewise not eliminated.

Overall, selection in the example given led to a yield of 78% correct constructs, whereas without selection only 41% of the constructs were correct. The result of the selection investigation is shown in FIG. 2.

EXAMPLE 2

The pFroup1 and 2 vectors illustrated in FIGS. 5-7 were synthesised, in which a nucleic acid to be investigated (insert) is linked with a reporter gene via a linker which comprises a translational coupler sequence. The vectors differ with regard to the nature of the translational coupling. In pFroup1 the stop and start consist of two separate codons (TAA ATG). In pFroup2 the stop codon overlaps with the start codon (TAATG). Any desired nucleic acid to be checked for frameshift mutations may be inserted at the position of the insert.

First of all, model inserts were purposefully tested to check the functionality of the vectors according to the invention. In doing so, a nucleic acid sequence coding for the green fluorescent protein (GFP), the reverse complementary sequence thereto (GFPrev) and part of the maltose binding protein (MBP) was inserted as insert into pFroup1 and 2. The GFP and GFPrev sequences have a correct reading frame, while the MBP insert does not have a sequence divisible by three and thus does not have a correct reading frame.

Functioning of the vectors was first of all demonstrated in a growth test. *E. coli* host cells containing the pFroup-GFP and pFroup-GFPrev vectors exhibited resistance to ampicillin and kanamycin. The pFroup-MBP vector does not impart kanamycin resistance to the host cell, as here, due to the incorrect reading frame, the Kan2 polypeptide is not formed due to premature termination of translation in the linker zone. In order to rule out the kanamycin resistance of the pFroup-GFP construct being due to the formation of a fusion protein from GFP and Kan2 (this could occur in pFroup1 bp skipping the stop codon), the synthesised GFP was investigated by immunoblot analysis (FIG. 8). It was possible to detect the free GFP, but not a very much larger GFP-Kan2 fusion protein. Correct functioning of translational coupling in the developed pFroup1 and 2 vectors was thus demonstrated.

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	acc Thr															480
	cgt Arg															528
	agc Ser								Gly							576
	gtg Val	_								-				-		624
	aac Asn 210															672
	agc Ser															720
	tat Tyr															768
_	ctg Leu	_		_					_	_	_	_	_		_	816
	gcg Ala															864
	cgt Arg 290															912
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	ggc Gly	_	_		_	_	_	_								1008
	ttt Phe								Val							1056
	acc Thr															1104
	acc Thr 370															1152
	acc Thr	_		_	-		_		_			_		_	_	1200
	acc Thr															1248
-	ggc Gly	_	_		_	_			Trp		_		_	-		1296
	ttt Phe															1344
ggc	gcg	gaa	ctg	ccg	ttt	acc	att	gat	aaa	taa	taa 🤉	ggc .	agc .	agc (cgt	1392

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gag Glu															
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Thr	Gly 50	Ser	Gln	His	Gln	Ala 55	Glu	Ser	Ile	Ser	Asn 60	Thr	Asn	Pro	Leu
Thr 65	Glu	Lys	Ala	Val	Ala 70	Ser	Val	Thr	Leu	Ala 75	Gly	Asn	Ser	Ser	Leu 80
CÀa	Pro	Ile	Arg	Gly 85	Trp	Ala	Val	His	Ser 90	Lys	Asp	Asn	Asn	Ile 95	Arg
Ile	Gly	Ser	Lys 100	Gly	Asp	Val	Phe	Val 105	Ile	Arg	Glu	Pro	Phe 110	Ile	Ser
CÀa	Ser	His 115	Leu	Glu	CAa	Arg	Thr 120	Phe	Phe	Leu	Thr	Gln 125	Gly	Ala	Leu
Leu	Asn 130	Asp	Lys	His	Ser	Asn 135	Gly	Thr	Val	Lys	Asp 140	Arg	Ser	Pro	His
Arg 145	Thr	Leu	Met	Ser	Cys 150	Pro	Val	Gly	Glu	Ala 155	Pro	Ser	Pro	Tyr	Asn 160
Ser	Arg	Phe	Glu	Ser 165	Val	Ala	Trp	Ser	Ala 170	Ser	Ala	СЛа	His	Asp 175	Gly
Thr	Ser	Trp	Leu 180	Thr	Ile	Gly	Ile	Ser 185	Gly	Pro	Asp	Asn	Gly 190	Ala	Val
Ala	Val	Leu 195	Lys	Tyr	Asn	Gly	Ile 200	Ile	Thr	Asp	Thr	Ile 205	Lys	Ser	Trp
Arg	Asn 210	Asn	Ile	Leu	Arg	Thr 215	Gln	Glu	Ser	Glu	Cys 220	Ala	Сув	Val	Asn
Gly 225	Ser	CÀa	Phe	Thr	Val 230	Met	Thr	Asp	Gly	Pro 235	Ser	Asn	Gly	Gln	Ala 240
Ser	Tyr	ГÀа	Ile	Phe 245	rys	Met	Glu	ГÀа	Gly 250	Lys	Val	Val	ГЛа	Ser 255	Val
Glu	Leu	Asp	Ala 260	Pro	Asn	Tyr	His	Tyr 265	Glu	Glu	CAa	Ser	Cys 270	Tyr	Pro
Asp	Ala	Gly 275	Glu	Ile	Thr	Cys	Val 280	Cys	Arg	Asp	Asn	Trp 285	His	Gly	Ser
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Tyr 305	Ile	Сув	Ser	Gly	Val 310	Phe	Gly	Asp	Asn	Pro 315	Arg	Pro	Asn	Asp	Gly 320
Thr	Gly	Ser	Cys	Gly 325	Pro	Met	Ser	Pro	Asn 330	Gly	Ala	Tyr	Gly	Val 335	Lys
Gly	Phe	Ser	Phe 340	Lys	Tyr	Gly	Asp	Gly 345	Val	Trp	Ile	Gly	Arg 350	Thr	Lys

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Ser	Thr	355	Ser	Arg	Ser	Gly	Phe 360	Glu	Met	Ile	Trp	Asp 365	Pro	Asn	Gly	
Trp	Thr 370	Gly	Thr	Asp	Ser	Ser 375	Phe	Ser	Val	Lys	Gln 380	Asp	Ile	Val	Ala	
Ile 385	Thr	Asp	Trp	Ser	Gly 390	Tyr	Ser	Gly	Ser	Phe 395	Val	Gln	His	Pro	Glu 400	
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		caq			aat	asa	aat	asa	+++	tta	ata	att	tta	<i>aa</i>	asa	48
		Gln			_	_	_	_			_	_			_	40
		atg Met			_	_		_				_		_		96
		gtt Val 35														144
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		gct Ala														336
		gcc Ala 115														384
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		ggt Gly					_		_	_			_	_	_	480
		gtt Val														528
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_	_			_		cac His 215						_				672		
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-	_	_			_	aat Asn		_	-			_		-	_	768		
	_	_	_	_	_	cag Gln	_	_						_	_	816		
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_		_			_	atc Ile 295	_	_	-		_	_				912		
_		_		_		gtt Val	_		_		_		_		_	960		
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Arg	Ala	Āla	Lys	Phe 245	Val	Asn	Asn	Ala	Ala 250	Gly	Phe	Leu	Phe	Ala 255	Āla		
	gtt Val															81	L6
	ttc Phe															86	54
	aat Asn 290															91	L2
	ttc Phe															96	50
	cac His															100	8 (
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	acg Thr															110	04
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	gct Ala															120	00
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	cgc Arg															129	96
	cag Gln															134	14
	ttt Phe 450															139	92
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acg	gca	ttc	ggt	gcc	cgc	ggt	cgc	gcc	aaa	ctg	cgc	gct	cgc	aat	cgg	153	36

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Thr	Ala	Phe	Gly 500	Ala	Arg	Gly	Arg	Ala 505	Lys	Leu	Arg	Ala	Arg 510	Asn	Arg		
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					cac His											1632	
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Pro	Phe	Val 35	His	Ala	Lys	Asn	Thr 40	Ile	Ala	Ile	Val	Ala 45	Ile	Phe	Ala		
Asn	Ala 50	Met	Gln	Phe	Asn	Arg 55	Ile	Phe	Pro	Leu	Phe 60	Gly	Gln	Ile	Phe		
Ile 65	Ala	Lys	Phe	His	Asn 70	Gly	Phe	Arg	Thr	Gln 75	Ile	Ile	Gln	Ala	Gln 80		
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		115			Thr		120			Ī		125					
	130				Gln	135					140						
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Phe	Phe	Val	Ile	Phe 165	Ala	Ala	Lys	Ile	Gln 170	Ala	Gly	Phe	Thr	Thr 175	Phe		
Asn	Leu	Gly	Thr 180	Arg	Asn	Gly	Ile	Phe 185	_	Lys	Ile	Phe	Thr 190	Gly	Arg		
Ala	Ala	Val 195	Val	Ile	His	Phe	Ala 200	Ala	Thr	Phe	Gln	Asn 205	Ile	Gly	Met		
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Ala 225	Phe	Gln	Lys	Gly	Ile 230	Phe	Ile	Phe	Phe	Val 235	Met	Phe	Ala	Ile	Phe 240		
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Lys Val Ala Ala Ala Gln Ala Thr Phe Phe Phe His Gln Ala 265 Val Phe Phe Ala Gly His Ala Gln Gly Phe Ala Ala Arg Ala Arg Ile Ala Asn Ala His Gly Thr Ile Gln Leu Ala Phe Leu Arg His Arg Phe Ala Phe Arg Ile Arg Phe Val Ala His Ala Phe Thr Phe Ala Ile Ala Leu His Lys Ala Ala Ala Ala Phe Thr Ala Ala Phe Phe Gly Phe Phe Asn Arg Asn Ala Phe Gln Asn Gly His Phe Ala Phe Arg Phe Thr His Phe Thr Lys Val His Ile Ile His Ala Phe Ile Phe Phe Gln Gln Met Gly Ala Gly Lys Val Ala Gln Ala Thr Ala Phe Leu Gln Phe Ala Ala Ala Ala Arg Arg His Lys His Ile Phe Phe Ala Phe Ala Thr Gly Thr Gln Val Pro Val Ala Val Val Ala Leu Val Leu Gln Met Ala Gly 410 Gln Arg Arg His Ala Ala Ile Phe Ile Thr Ala Phe Asn Ala Thr Phe 425 Ala Gln His Gly Lys Ile Lys Ile Phe Arg Phe Gln Ala Gly Phe Ala 440 Val Phe Met Gln Ile Val Arg Gln Ala Ala Phe Ala Thr Lys Phe Gln Ala Leu Arg Ala Asn Lys Ile His Ile Lys Thr Ala Arg Gly Phe Ala 475 Lys Gln Thr Leu Phe His His Thr Phe Pro Ala Ala Arg His Ala Phe 490 Thr Ala Phe Gly Ala Arg Gly Arg Ala Lys Leu Arg Ala Arg Asn Arg 505 Phe Leu Gln Gln Arg Val Ala Phe Ala Ala Thr Phe Asn His His Leu Phe Phe Ile Phe Ala His Thr Phe Gln Phe His Arg His Ile Phe Phe 535 Thr Gln Ile Val Arg Arg Ile Arg His Gln Thr Phe Ala Ala Gly His Gly Arg Arg Arg Ala Thr Ile Ala Gln Ala Gln Ala Phe Gln Ala Gly Gly Ala His Gly Arg Gln Phe Gly Thr <210> SEQ ID NO 11 <211> LENGTH: 1755 <212> TYPE: DNA <213 > ORGANISM: Homo sapiens <220> FEATURE: <221> NAME/KEY: CDS <222> LOCATION: (1)..(1740) <400> SEQUENCE: 11 ggt acc aaa ctg gcg acc atg ggc acc acc ggc ctg gaa agc ctg agc Gly Thr Lys Leu Ala Thr Met Gly Thr Thr Gly Leu Glu Ser Leu Ser 5

ctg ggc gat cgt ggc gcg gcg ccg acc gtg acc agc agc gaa cgt ctg

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										cag Gln							192
S										ccg Pro							240
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G	lu	Arg	Val	Val 100	Lys	Gln	Arg	Leu	Phe 105	ggc Gly	Lys	Thr	Ala	Gly 110	Arg	Phe	336
_						_	_	_		ctg Leu			_	_		_	384
S	er	Leu 130	Ala	Asn	Tyr	Leu	His 135	Lys	Asn	ggc Gly	Glu	Thr 140	Ser	Leu	Lys	Pro	432
G 1	1u 45	Asp	Phe	Asp	Phe	Thr 150	Val	Leu	Ser	aaa Lys	Arg 155	Gly	Ile	ГÀа	Ser	Arg 160	480
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										ctg Leu							672
G 2	1y 25	Val	Asp	Asp	Val	Asn 230	Phe	Arg	Lys	gtg Val	Arg 235	Lys	Pro	Lys	Gly	Lys 240	720
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А	rg	Lys	Ser	Cys 260	Ser	Gly	Phe	Val	Gln 265	agc Ser	Asp	Ser	ГÀв	Arg 270	Glu	Ser	816
										gaa Glu							864
	_	_	_	_			_		_	gat Asp				_		_	912
Т		_	_			_	_	-		agc Ser	_					_	960
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					ttt Phe											1104	
					aaa Lys											1152	
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					ctg Leu											1632	
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Val Pro Asp Pro Pro Asn Asp Leu Arg Lys Glu Asp Val Ala Met Glu

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Ser	Asn	Asn	Ser 180	Asn	Trp	Asn	Leu	Arg 185	Thr	Arg	Ser	ГÀа	Cys	Lys	ГЛа
Asp	Val	Phe 195	Met	Pro	Pro	Ser	Ser 200	Ser	Ser	Glu	Leu	Gln 205	Glu	Ser	Arg
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Val	Thr	Ile	Leu	Lys 245	Gly	Ile	Pro	Ile	Lys 250	Lys	Thr	Lys	Lys	Gly 255	Cys
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Val	Cys	Asn 275	Lys	Ala	Asp	Ala	Glu 280	Ser	Glu	Pro	Val	Ala 285	Gln	Lys	Ser
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Glu	Asp	Thr	Ile	Pro 405	Arg	Thr	Gln	Ile	Glu 410	Arg	Arg	Гла	Thr	Ser 415	Leu
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Leu	Ser	Leu	Ser 580													
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	2> T			7		-1 6	·									
)> FE			ALUI	lfici	lai s	eque	ence								
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	_	_		_	cat His	_		_	_		_				918
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Val					aaa Lys										1062
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					cag Gln									-	1254
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gat atc cat atg gaa ttc gtc gac aag ctt cta gat ctg gct agc acc
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Asp Ile His Met Glu Phe Val Asp Lys Leu Leu Asp Leu Ala Ser Thr
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Ala Pro Glu Leu Phe Leu Lys His Gly Lys Gly Ser Val Ala Asn Asp
Val Thr Asp Glu Met Val Arg Leu Asn Trp Leu Thr Glu Phe Met Pro
Leu Pro Thr Ile Lys His Phe Ile Arg Thr Pro Asp Asp Ala Trp Leu
Leu Thr Thr Ala Ile Pro Gly Lys Thr Ala Phe Gln Val Leu Glu Glu
Tyr Pro Asp Ser Gly Glu Asn Ile Val Asp Ala Leu Ala Val Phe Leu
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Arg Arg Leu His Ser Ile Pro Val Cys Asn Cys Pro Phe Asn Ser Asp
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 \hbox{Arg Val Phe Arg Leu Ala Gln Ala Gln Ser Arg Met Asn Asn Gly Leu} \\
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Val Asp Ala Ser Asp Phe Asp Asp Glu Arg Asn Gly Trp Pro Val Glu
Gln Val Trp Lys Glu Met His Lys Leu Leu Pro Phe Ser Pro Asp Ser
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Val Val Thr His Gly Asp Phe Ser Leu Asp Asn Leu Ile Phe Asp Glu
Gly Lys Leu Ile Gly Cys Ile Asp Val Gly Arg Val Gly Ile Ala Asp
Arg Tyr Gln Asp Leu Ala Ile Leu Trp Asn Cys Leu Gly Glu Phe Ser
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1 5
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The invention claimed is:

- 1. A method for identifying frameshift mutations in coding target nucleic acids comprising one or more stop codons which comprises the steps:
 - (i) providing a host cell comprising a double-stranded 20 nucleic acid, which comprises a coding target nucleic acid and a coding opposite strand nucleic acid complementary thereto, in which the opposite strand nucleic acid is linked via a linker with a reporter gene in 3'-position; wherein the linker comprises a translational coupler sequence which comprises a stop codon in frame to the reading frame of the opposite strand nucleic acid and a start codon.
 - wherein the reporter gene is in frame to the start codon wherein the linker comprises further stop codons in reading frames shifted by +1 and -1,
 - wherein the further stop codons are located upstream of the stop codon of the translational coupler sequence,
 - wherein the distance from the further stop codon to the start codon of the translational coupler sequence is at least 30 base pairs, such that no translational coupling occurs.
 - (ii) effecting expression of the opposite strand nucleic acid; and
 - (iii) identifying whether expression of the reporter gene occurs in the host cell, in which expression of the reporter gene indicates that the target nucleic acid does not comprise a frameshift mutation.

- 2. A method according to claim 1, wherein the start and stop codons of the translational coupler sequence follow on immediately from one another.
- 3. A method according to claim 1, wherein the start and stop codons of the translational coupler sequence overlap with one another.
- **4**. A method according to claim **1**, wherein the start and stop codons of the translational coupler sequence are at a distance from one another which is selected such that translational coupling is enabled.
- **5**. A method according to claim **1**, in which the opposite strand nucleic acid is furthermore in operative linkage with an expression control sequence in 5'-position.
- **6**. A method according to claim **1**, in which step (i) comprises: introducing an expression vector comprising the double stranded nucleic acid into a host cell.
- 7. A method according to claim 6, in which the opposite strand nucleic acid in the expression vector is in operative linkage with an expression control sequence in 5'-position.
- **8**. A method according to claim **6**, in which an expression vector is used in step (i) which additionally comprises at least one selection marker gene.
- **9**. A method according to claim **8**, in which the selection marker gene is constitutively expressed.
- 10. A method according to claim 6, in which the expression vector is introduced by calcium phosphate coprecipitation, lipofection, electroporation, particle bombardment or viral infection.

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